Ezrin enrichment on curved cell membranes requires phosphorylation or interaction with a curvature-sensitive partner

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Proper spatial localisation of proteins that connect cortical actin and the plasma membrane is essential for cell shaping and function. *In vivo*, the ERM protein ezrin is associated on membranes that are flat, or with positive or negative curvature. To assess the curvature-sensing of ezrin, we use cell biology and *in vitro* approaches that associate cryo-electron microscopy (cryo-EM) and mechanical measurements on model membranes containing PIP₂ and purified ezrin. We observe that ezrin (ezrinWT) and its constitutively phosphorylated mutant (ezrinTD) self-assemble in an anti-parallel manner, zipping adjacent membranes. Phosphorylation reduces ezrin intermolecular interactions, induces a conformational change, and facilitates ezrin binding to actin filaments, as shown by cryo-EM, and promotes ezrin binding to positively curved membrane. While neither ezrinTD nor ezrinWT senses negative membranes such as protrusions requires them to interact with curvature sensors, e.g. I-BAR-domain proteins. Overall, our work corroborates a role for ezrin, not as a curvature sensor but rather in the mechanical cohesion of membranes with actin.



Figure 1. (A) Representative cryo-electron micrograph of PIP₂-membranes tethered by ezrinTD. Scale bar, 20 nm. (B) Representative cryo-electron micrograph of PIP₂-containing LUVs incubated with muscle F-actin in the presence of ezrinTD. Arrowheads indicate F-actin and arrows indicate tethered vesicles. (C) Representative structured illumination microscopy images of cellular protrusions of LLC-PK1 cells transfected with GFP-I-BAR domain and immunolabeled for endogenous ezrin. Scale bar, 2 μ m (D) (Top) Representative confocal images of ezrinTD in IRSp53 I-BAR domain induced tubules. Scale bar, 5 μ m (Bottom) Normalized fluorescence intensity profiles along the line drawn from outside the GUV towards the interior of the GUV, as indicated in the top image.